

## THE EFFECTS OF ACTIVATED CHLORINE DIOXIDE (ClO<sub>2</sub>) ON SELECTED BACTERIAL PURE CULTURES

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**Abstract.** A series of experiments were done to determine the effects of activated chlorine dioxide (ClO<sub>2</sub>) on selected bacterial pure cultures related with food-borne diseases and those that serve as indicators of proper hygiene and handling in seafood, particularly black tiger shrimps. These bacterial cultures were *Escherichia coli* (ATCC 11775), *Salmonella typhimurium* (ATCC 14028), *Staphylococcus aureus* (ATCC 25923), as well as three *Vibrios*: *V. parahaemolyticus* (AQ3815, Kyoto University, Japan), *V. cholerae* and *V. vulnificus* taken from the Universiti Kebangsaan Malaysia's pure culture collection. Pure cultures were exposed to five ClO<sub>2</sub> concentrations (20, 40, 60, 80, and 100 parts per million (ppm)) for 15, 30 and 45 min. Zero ppm ClO<sub>2</sub> at 0 min exposure served as the control. Activated ClO<sub>2</sub> was found successful in reducing bacterial population. At 20 ppm, the halophilic *V. parahaemolyticus* and *V. vulnificus* were eliminated, while *V. cholerae*, was eliminated only at 60 ppm. *E. coli* and *S. typhimurium* were totally eliminated after 30 min of exposure with 100 ppm activated ClO<sub>2</sub> while *V. cholerae* was eliminated after 30 min at 20 and 40 ppm ClO<sub>2</sub>. Further studies will involve spiking bacterial cultures into tiger shrimps and other sea foods to see whether ClO<sub>2</sub> can exhibit the same reducing action on bacterial population in a food matrix.

**Abstrak.** Suatu siri kajian telah dijalankan bagi menentukan kesan klorin dioksida teraktif (ClO<sub>2</sub>) terhadap kultur bakteria terpilih yang berkaitan dengan penyakit bawaan makanan serta bertindak sebagai penunjuk kebersihan semasa mengendalikan makanan laut, terutamanya udang harimau. Kulture bakteria tersebut adalah *Escherichia coli* (ATCC 11775), *Salmonella typhimurium* (ATCC 14028), *Staphylococcus aureus* (ATCC 25923), serta tiga sepsis *Vibrios*: *V. parahaemolyticus* (AQ3815, Kyoto University, Japan), *V. cholerae* and *V. vulnificus* yang diambil daripada kultur simpanan Universiti Kebangsaan Malaysia. Kultur tulen tersebut telah didedahkan kepada lima kepekatan ClO<sub>2</sub> (20, 40, 60, 80, dan 100 bahagian per juta (ppm)) selama 15, 30 dan 45 min. Sifar ppm ClO<sub>2</sub> pada pendedahan 0 min bertindak sebagai kawalan. ClO<sub>2</sub> teraktif didapati berjaya mengurangkan populasi bakteria tersebut. Pada 20 ppm, *V. parahaemolyticus* dan *V. vulnificus* yang halofilik telah dimusnahkan, manakala *V. cholerae*, telah dimusnahkan hanya pada 60 ppm. *E. coli* dan *S. typhimurium* dimusnahkan selepas 30 min pendedahan dengan 100 ppm ClO<sub>2</sub> teraktif, manakala *V. cholerae* dimusnahkan selepas 30 min pendedahan pada 20 dan 40 ppm ClO<sub>2</sub>. Kajian selanjutnya akan melibatkan penginokulan kultur-kultur bakteria ke dalam udang harimau bagi menentukan samada ClO<sub>2</sub> berupaya memberi tindakan pengurangan terhadap populasi bakteria di dalam suatu matrik makanan.

**Keywords:** Chlorine dioxide; *E.coli*; *S. typhimurium*; *S. aureus*; *V. cholerae*; *V. parahaemolyticus*; *V. vulnificus*

### Introduction

The risk of foodborne illness has increased markedly over the last 20 years, with nearly a quarter of the population at higher risk for illness today. Consequently, preventing foodborne illness and death remains a major public health challenge [1].

*Escherichia coli* and *Salmonella typhimurium* are classified as organisms causing food-borne infections while *Staphylococcus aureus* is considered one that causes food-borne intoxication [2]. Roberts and Greenwood [2] further state that *E. coli* is a microorganism that can be used as a marker to demonstrate that faecal pollution may have occurred at some stage during the production of food. The virulence of *S. typhimurium* grown in different foods varies. The possibility exists, therefore, that ingestion of foods containing the same number of salmonellae may not constitute a similar health hazard [3].

Foods that require considerable handling during preparation and that are kept at slightly elevated temperatures after preparation are frequently involved in staphylococcal food poisoning [4]. This microorganism is frequently introduced into food by food handlers and indirectly by equipment [2] and can therefore be considered indicative of proper food handling and hygiene.

Strains of the genus *Vibrio* are ubiquitous in the estuarine environment and as might be expected, given their environmental distribution, vibrios are a frequent isolate from seafood [5]. Only a few have been found to be responsible for food-borne infection, and of these, *V. cholerae*, *V. parahaemolyticus*, and *V. vulnificus* occur most frequently [6].

Chlorine dioxide (ClO<sub>2</sub>), is a chemical compound which has been discovered to possess great potential in reducing microbial load. Junli et al. [7] confirms that the killing effect of ClO<sub>2</sub> on bacteria is similar to or better than that of liquid chlorine, at a relatively wider pH range, making it an excellent disinfectant substitute. Chlorine dioxide is registered with the Environmental Protection Agency (EPA Registration No.74986-1) [8], and is considered an excellent bactericide, fungicide and antimicrobial agent. The EPA also confirms that a post oxidant dose of 0.2-1 mg/l and a residual of 0.1-0.5 mg/l of ClO<sub>2</sub> effectively inactivate pathogenic viruses, bacteria and protozoa. The use of chlorine dioxide on food has been approved by the Food and Drug Administration [9].

Information on the utilization of ClO<sub>2</sub> as a bactericide in seafood products is limited [10]. This study hopes to provide technical data that may, in the future, be useful as basis for the use of ClO<sub>2</sub> on different food and food systems. The objective of this study was to determine the effectivity of different ClO<sub>2</sub> concentrations at different exposure times in reducing pure culture populations of the above mentioned pathogens.

## Experimental

### *Preparation of Working Cultures*

Working cultures of *E. coli*, *S. typhimurium*, *S. aureus*, and *V. cholerae* (Vc) were prepared by streaking a loopful onto Nutrient Agar (Oxoid). Working cultures of *V. parahaemolyticus* and *V. vulnificus* were prepared by streaking a loopful onto NA with 3% NaCl (NA+3% S) content. All NA and NA+3% S plates were then incubated overnight at 37°C.

After incubation, individual colonies of different cultures were inoculated into Tryptic Soya Broth (TSB). *V. parahaemolyticus* and *V. cholerae* were inoculated into TSB added with 3% salt (TSB+3% S). TSB and TSB+3% S were then incubated overnight at 37°C.

### *Preparation and Activation of Chlorine Dioxide*

Chlorine dioxide (Aqua 5) was obtained from Mitrol Technology Pte Ltd. The liquid stock solution had a 50,000 ppm concentration. Chlorine dioxide was prepared in glass bottles in an open area. The needed concentrations of ClO<sub>2</sub> were prepared based on the following equation:

$$c_1v_1=c_2v_2,$$

where,

c<sub>1</sub> = Concentration of stock solution (Aqua 5) = 50,000 ppm

v<sub>1</sub> = Volume ClO<sub>2</sub> needed to be prepared

c<sub>2</sub> = Target concentration of Stabilized ClO<sub>2</sub> (20, 40, 60, 80, 100 ppm)

v<sub>2</sub> = Target volume of distilled water

Chlorine dioxide was activated using food-grade citric acid. The amount of citric acid used was 10% of the amount of stock solution needed to prepare the different concentrations of ClO<sub>2</sub>. A sample computation of both preparation and activation of ClO<sub>2</sub> is provided below:

E.g. To prepare 100 ml of 100 ppm ClO<sub>2</sub>

$$v_1 = c_2v_2/c_1,$$

$$v_1 = (100 \text{ ppm})(100 \text{ ml})/ 50,000 \text{ ppm}$$

$$v_1 = 0.2 \text{ ml ClO}_2 \text{ needed}$$

To activate, we get 10% of the amount of needed ClO<sub>2</sub>:

$$(0.2 \text{ ml})(0.1) = 0.02 \text{ g of citric acid for activation}$$

Therefore, 0.02 g of citric acid is to be added to 0.2 ml ClO<sub>2</sub> in order to prepare 100 ml of 100 ppm ClO<sub>2</sub> solution.

Citric acid was added to the  $\text{ClO}_2$  and left to activate for 10-15 minutes or until a yellow color developed and gas evolved. The glass container was not covered to allow the escape of gas, a result indicative of the completion of the chemical reaction. The appropriate amount of water was then added. Preparation of  $\text{ClO}_2$  concentrations was done right before application.

#### Application of Chlorine Dioxide

Prepared bacterial cultures inoculated in 10 ml portions of TSB and TSB+3% S were transferred aseptically into individual 15 ml-capacity centrifuge tubes and centrifuged at 3600 rpm at 27°C for 15 min to obtain cell pellets with clear supernatant. The supernatant was then decanted and, in place, 10 ml of freshly prepared activated  $\text{ClO}_2$  was poured into the centrifuge tube, capped tightly, vortexed to allow dispersal of the cultures from its concentrated state, and kept for the desired amount of time.

#### Enumeration of Bacteria

After exposure to  $\text{ClO}_2$  at different times, the surface-colony drop-plate method [2] on Plate Count Agar (PCA) was used for counting *E. coli*, *S. typhimurium*, and *S. aureus*. Plate Count Agar is a non-selective medium used for general viable counts of bacteria in food [11]. *Vibrio cholerae*, *V. parahaemolyticus*, and *V. vulnificus* were enumerated using the spread plate method [12]. For *V. cholerae*, enumeration was done on thiosulfate-citrate bile salts sucrose agar (TCBS) while *V. parahaemolyticus* and *V. vulnificus* were enumerated on TCBS+3% S because of their halophilic nature. TCBS agar is a selective isolation medium for pathogenic vibrios [13]. The selective agents in the media are Sodium thiosulphate, sodium taurocholate and ferric citrate [6], making it suitable for growth of *V. cholerae*, *V. parahaemolyticus* and other vibrios [14]. A control sample (0 ppm or 0 min) was employed. All experiments were done in duplicate.

#### Statistical Analysis

The statistical analysis, one-way Analysis of Variance (ANOVA), with Turkey's family error rate comparison, was done using the MINITAB Release 12.1.

### Results And Discussion

#### The Effect of Different $\text{ClO}_2$ Concentrations

Figure 1 shows the effects of various activated  $\text{ClO}_2$  concentrations when applied to bacterial pure cultures for 15 min. A decreasing trend of colony counts as  $\text{ClO}_2$  concentrations increase was seen. The effectivity of most  $\text{ClO}_2$  concentrations, as per bacterial culture, significantly differs from one another (see Table 1). At 20 ppm, the halophilic *V. parahaemolyticus* and *V. vulnificus* were eliminated, while *V. cholerae*, was eliminated only at 60 ppm.

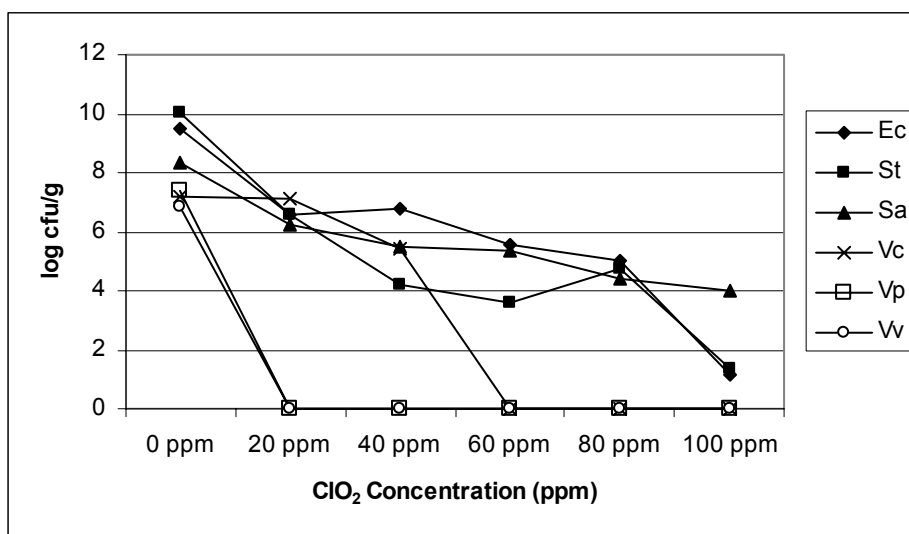


FIGURE 1. The Effect of  $\text{ClO}_2$  on Bacterial Pure Cultures after 15 Min Exposure Time

The ability of ClO<sub>2</sub> to kill bacteria was studied by Junli et al. [7], who after finding similar results, stated that the bactericidal effect of ClO<sub>2</sub> is due to its 100 % existence in molecular form when dissolved in water. Further, Junli's study indicated that because of this molecular state, ClO<sub>2</sub> easily permeated through cell membranes and into the bacterial body, thus exhibiting its bactericidal effect. *E. coli*, *S. typhimurium*, and *S. aureus* were not eliminated even with treatments of up to 100 ppm at 15 min exposure time.

TABLE 1. Effects of ClO<sub>2</sub> on Log Reduction of Bacterial Pure Cultures at 15 Min Exposure Time

Pure Culture	Log cfu/g					
	0 ppm	20 ppm	40 ppm	60 ppm	80 ppm	100 ppm
Ec	9.52 <sup>*a</sup>	6.59 <sup>b</sup>	6.75 <sup>c</sup>	5.59 <sup>d</sup>	5.01 <sup>e</sup>	1.19 <sup>f</sup>
St	10.06 <sup>a</sup>	5.58 <sup>b</sup>	4.20 <sup>c</sup>	3.62 <sup>d</sup>	4.78 <sup>e</sup>	1.37 <sup>f</sup>
Sa	8.36 <sup>a</sup>	6.22 <sup>b</sup>	5.46 <sup>bc</sup>	5.36 <sup>c</sup>	4.43 <sup>d</sup>	4.01 <sup>e</sup>
Vc	7.19 <sup>a</sup>	7.10 <sup>a</sup>	5.39 <sup>b</sup>	0.00 <sup>c</sup>	0.00 <sup>c</sup>	0.00 <sup>c</sup>
Vp	7.37 <sup>a</sup>	0.00 <sup>b</sup>	0.00 <sup>b</sup>	0.00 <sup>b</sup>	0.00 <sup>b</sup>	0.00 <sup>b</sup>
Vv	6.83 <sup>a</sup>	0.00 <sup>b</sup>	0.00 <sup>b</sup>	0.00 <sup>b</sup>	0.00 <sup>b</sup>	0.00 <sup>b</sup>

\*Results are the mean scores of 2 replications

<sup>a-f</sup> Means with two different lowercase letter superscripts for rows are significantly different (p<0.05)

*The Effect of Extended Exposure Times*

Figure 2 illustrates the result of extended exposure time (30 and 45 min) on colonies of bacterial pure cultures. *E. coli* and *S. typhimurium* were totally eliminated after 30 min of exposure with 100 ppm activated ClO<sub>2</sub> while *V. cholerae* was eliminated after 30 min at 20 and 40 ppm ClO<sub>2</sub>. *S. aureus* was not eliminated even after 30 min of exposure with activated 100 ppm ClO<sub>2</sub>. This may be due to the structure of *S. aureus*; it is gram-positive and forms a cross, three-dimensional spatial network structure of large mechanical strength in four-peptide side chain which is difficultly contacted by disinfectants, making it stronger than gram-negative bacteria [7]. *S. aureus* was totally eliminated after 45 min exposure with 100 ppm activated ClO<sub>2</sub>.

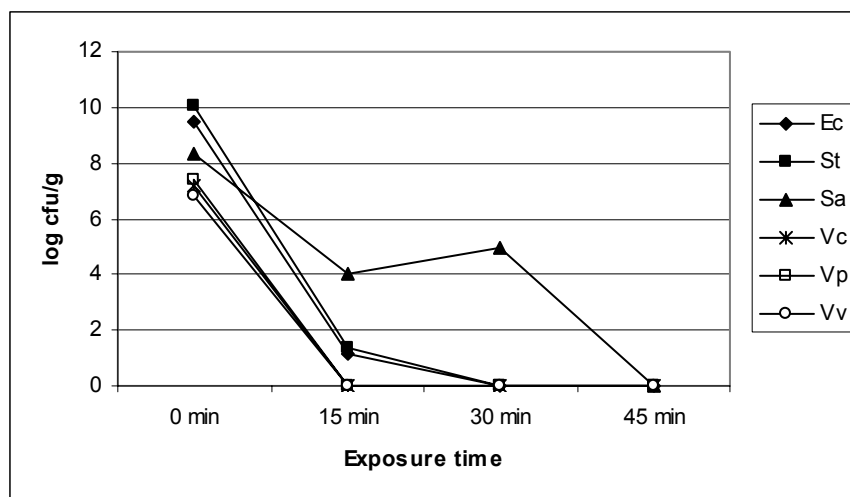


FIGURE 2. The Effect of Prolonged ClO<sub>2</sub> (100 ppm) Exposure Time on Bacterial Pure Cultures

Table 2 tabulates the effects of prolonged ClO<sub>2</sub>-exposure on microbial population. No reduction trend was seen for the different exposure times. Although the number of microbial colonies does decrease as ClO<sub>2</sub>-exposure time increases, not all these changes were significantly different from one another. Previous studies by Du et al. [15] on the efficacy of ClO<sub>2</sub> gas in reducing *E. coli* O157:H7 on apple surfaces suggested comparable results where the log reductions increase with the increase of treatment time at the 3.3-7.2 mg/l levels, as did similar studies by Han et al. [16] on green pepper surfaces. It is also noted that *S. aureus* was the microorganism least affected by prolonged ClO<sub>2</sub> exposure.

TABLE 2. Effects of ClO<sub>2</sub> at 15, 30 and 45 Min Exposure Times on Bacterial Pure Cultures

Microbes/ exposure time	Log cfu/g					
	0 ppm	20 ppm	40 ppm	60 ppm	80 ppm	100 ppm
<i>E. coli</i>						
15 min	9.52 <sup>*Aa</sup>	6.59 <sup>Ab</sup>	6.75 <sup>Ac</sup>	5.59 <sup>Ad</sup>	5.01 <sup>Ae</sup>	1.19 <sup>Af</sup>
30 min	9.52 <sup>Aa</sup>	6.63 <sup>Ab</sup>	5.77 <sup>Bc</sup>	5.72 <sup>Ac</sup>	0.49 <sup>Ad</sup>	0.00 <sup>Be</sup>
45 min	9.52 <sup>Aa</sup>	6.49 <sup>Ab</sup>	5.65 <sup>Bc</sup>	2.47 <sup>Bd</sup>	1.38 <sup>Ae</sup>	0.00 <sup>Bf</sup>
<i>S. typhimurium</i>						
15 min	10.06 <sup>Aa</sup>	6.58 <sup>Ab</sup>	4.20 <sup>Ac</sup>	3.62 <sup>Ad</sup>	4.78 <sup>Ae</sup>	1.37 <sup>Af</sup>
30 min	10.06 <sup>Aa</sup>	5.45 <sup>Bab</sup>	5.42 <sup>Bab</sup>	1.14 <sup>Bb</sup>	1.44 <sup>Bb</sup>	0.00 <sup>Bc</sup>
45 min	10.06 <sup>Aa</sup>	6.35 <sup>Ab</sup>	2.27 <sup>Cc</sup>	1.11 <sup>Bd</sup>	0.00 <sup>Cc</sup>	0.00 <sup>Be</sup>
<i>S. aureus</i>						
15 min	8.36 <sup>Aa</sup>	6.22 <sup>Ab</sup>	5.46 <sup>Abc</sup>	5.36 <sup>Ac</sup>	4.43 <sup>Ad</sup>	4.01 <sup>Ae</sup>
30 min	8.36 <sup>Aa</sup>	5.53 <sup>Abc</sup>	4.31 <sup>Bbc</sup>	4.83 <sup>Abc</sup>	4.20 <sup>Ac</sup>	2.94 <sup>Ad</sup>
45 min	8.36 <sup>Aa</sup>	4.65 <sup>Ab</sup>	3.87 <sup>Cb</sup>	3.77 <sup>Ab</sup>	3.68 <sup>Ab</sup>	0.00 <sup>Ac</sup>
<i>V. cholerae</i>						
15 min	7.19 <sup>Aa</sup>	7.10 <sup>Aa</sup>	5.39 <sup>Ab</sup>	0.00 <sup>Ac</sup>	0.00 <sup>Ac</sup>	0.00 <sup>Ac</sup>
30 min	7.19 <sup>Aa</sup>	0.00 <sup>Bb</sup>	0.00 <sup>Bb</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>
45 min	7.19 <sup>Aa</sup>	0.00 <sup>Bb</sup>	0.00 <sup>Bb</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>
<i>V. parahemolyticus</i>						
15 min	7.37 <sup>Aa</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>
30 min	7.37 <sup>Aa</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>
45 min	7.37 <sup>Aa</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>
<i>V. vulnificus</i>						
15 min	6.83 <sup>Aa</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>
30 min	6.83 <sup>Aa</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>
45 min	6.83 <sup>Aa</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>	0.00 <sup>Ab</sup>

\* Results are the mean scores of 2 replications.

<sup>A-C</sup> Means with two different uppercase letter superscripts for columns and each bacterial culture are significantly different (p<0.05).

<sup>a-c</sup> Means with two different lowercase letter superscripts for rows and each bacterial culture are significantly different (p<0.05).

### Conclusions

From the experiment conducted, the following can be concluded:

- As ClO<sub>2</sub> concentration increases, microbial population decreases.
- Different ClO<sub>2</sub> concentrations (0, 20, 40, 60, 80, and 100 ppm) have significantly different reducing effects on bacterial pure cultures.
- Longer exposure of pure cultures to ClO<sub>2</sub> further reduces colony counts but the differences observed were not significantly different.
- Activated ClO<sub>2</sub> is effective in reducing and eliminating certain microbial pure cultures, especially in halophilic *Vibrios* even at relatively low concentrations.

In continuation of this study, the effect of activated ClO<sub>2</sub> in reducing and ultimately eliminating microbiological population in food is being examined. The food of interest is black tiger shrimps (*Penaeus monodon*).

Experiments are being conducted to determine whether or not activated ClO<sub>2</sub> would exhibit the similar microbial-reducing actions when applied to a food matrix. The success of above mentioned will provide technical information that will further support the use of ClO<sub>2</sub> in the food, and particularly the seafood industry as an alternative anti-microbial agent, so as to meet safety measures dictated by consumers and the world market in general.

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